

Mechano-Hypoxia Conditioning of Human Stem Cell-Based Cartilage with Transient TGF-β3 Stimulation

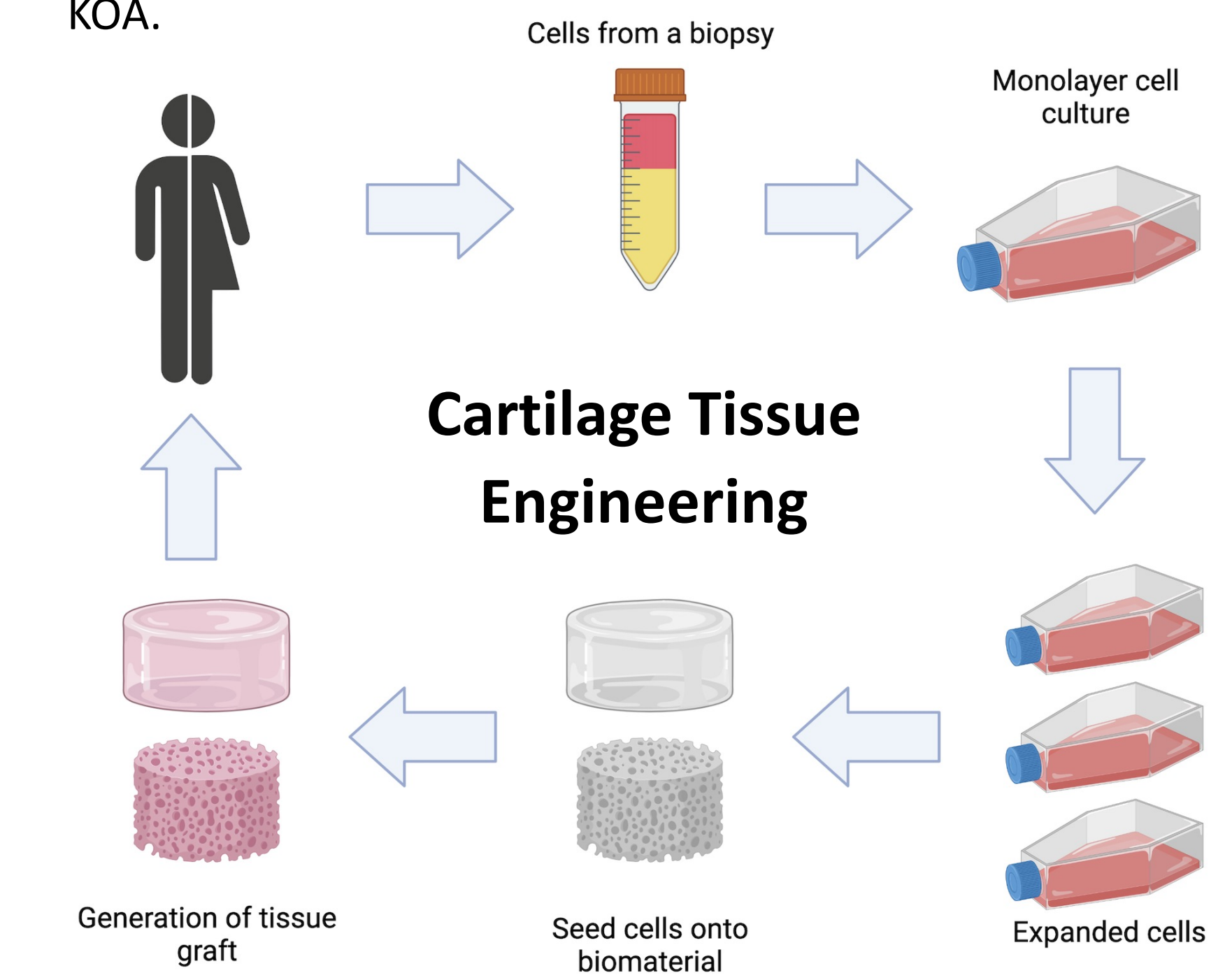
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Introduction

- Clinical Need**
- Knee osteoarthritis (KOA) affects > 650M people globally.
 - Injured articular cartilage (AC) increase the risk of KOA.
 - AC is avascular and has a limited self-repair capacity, making injuries difficult to treat.
 - Cartilage tissue engineering using cell-based methods aims to generate functional replacements to treat cartilage defects and KOA.



- Cell Selection**
- Human bone marrow mesenchymal stem cells (hBM-MSC) can differentiate into chondrocytes.
 - However, they are prone to hypertrophic differentiation upon stimulation with growth factors like TGF-β1/3.

- Mechano-Hypoxia Conditioning**
- Simulates physiologically relevant conditions of the knee joint (mechanical loading and hypoxia)
 - Mechano-hypoxia conditioning of engineered meniscus [1]:
 - ✓ increased the gene expression of hyaline cartilage markers, *SOX9* and *COL2A1*.
 - ✓ inhibited hypertrophic marker *COL10A1*.
 - ✓ promoted mechanical property development.

Hypothesis

We hypothesize that **mechano-hypoxia conditioning** will promote **non-hypertrophic** differentiation of **hBM-MSC** embedded in an **hyaluronan (HA)** hydrogel to **chondrocytes** with **transient TGF-β3** stimulation.

1. Szojka et al. (2021) Mechano-Hypoxia Conditioning of Engineered Human Meniscus. Front. Bioeng. Biotechnol. 9:739438. doi: 10.3389/fbioe.2021.739438

Methods

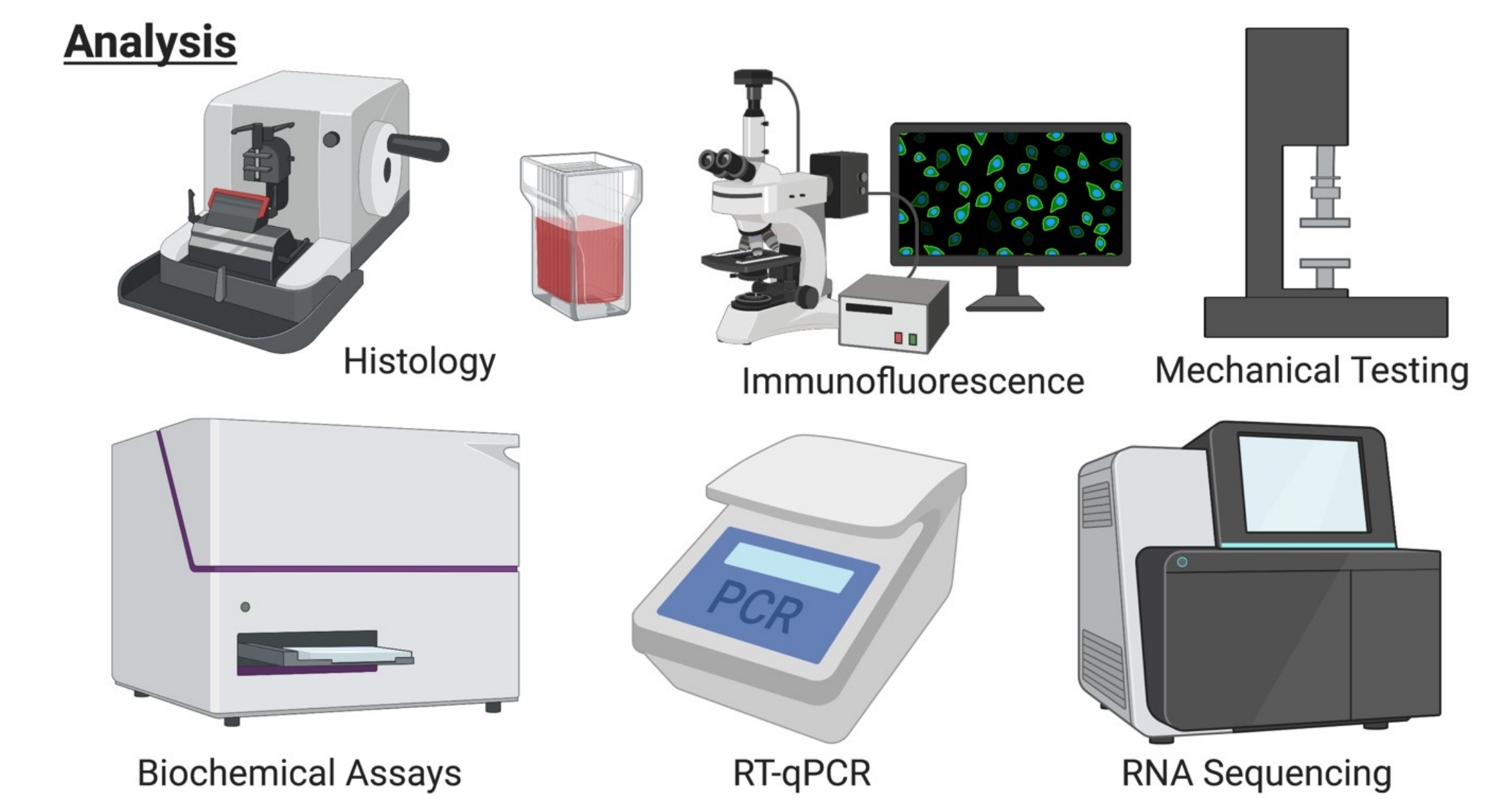
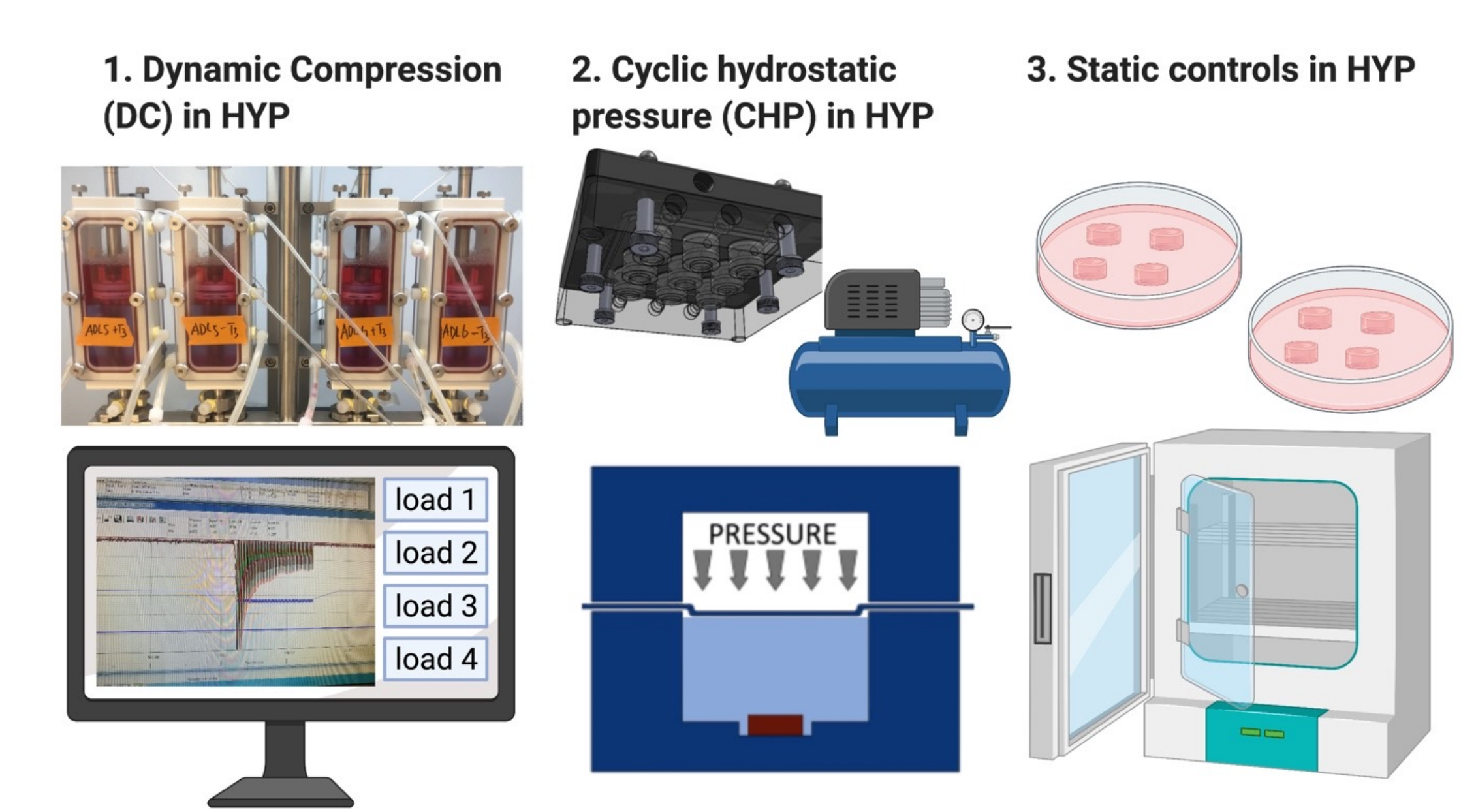
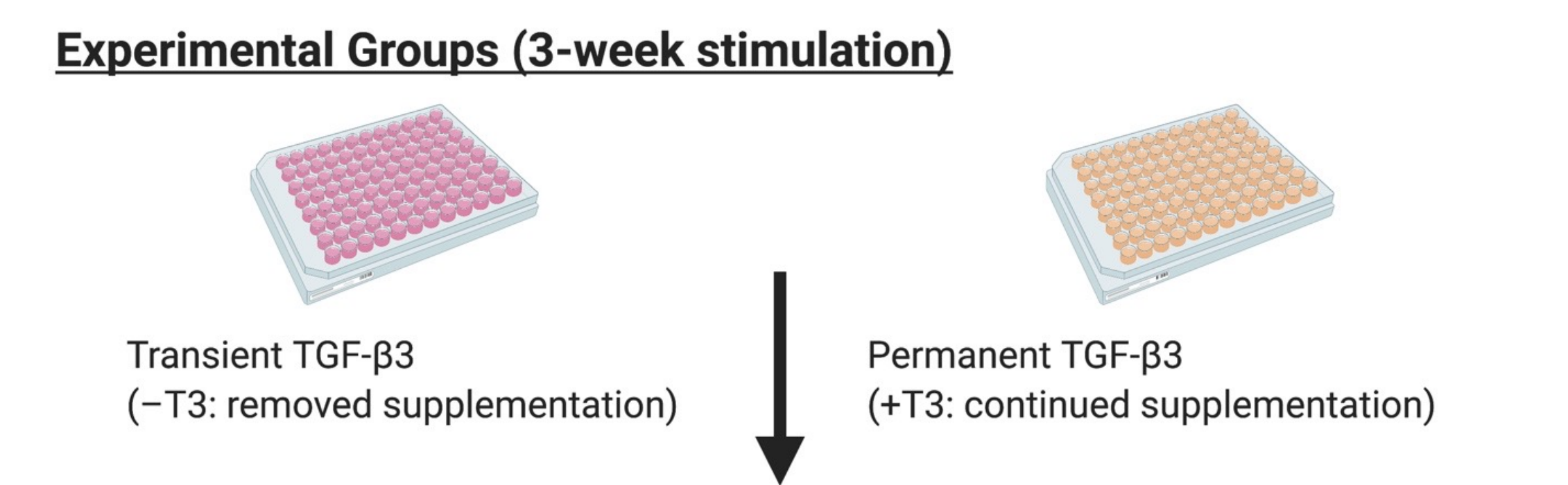
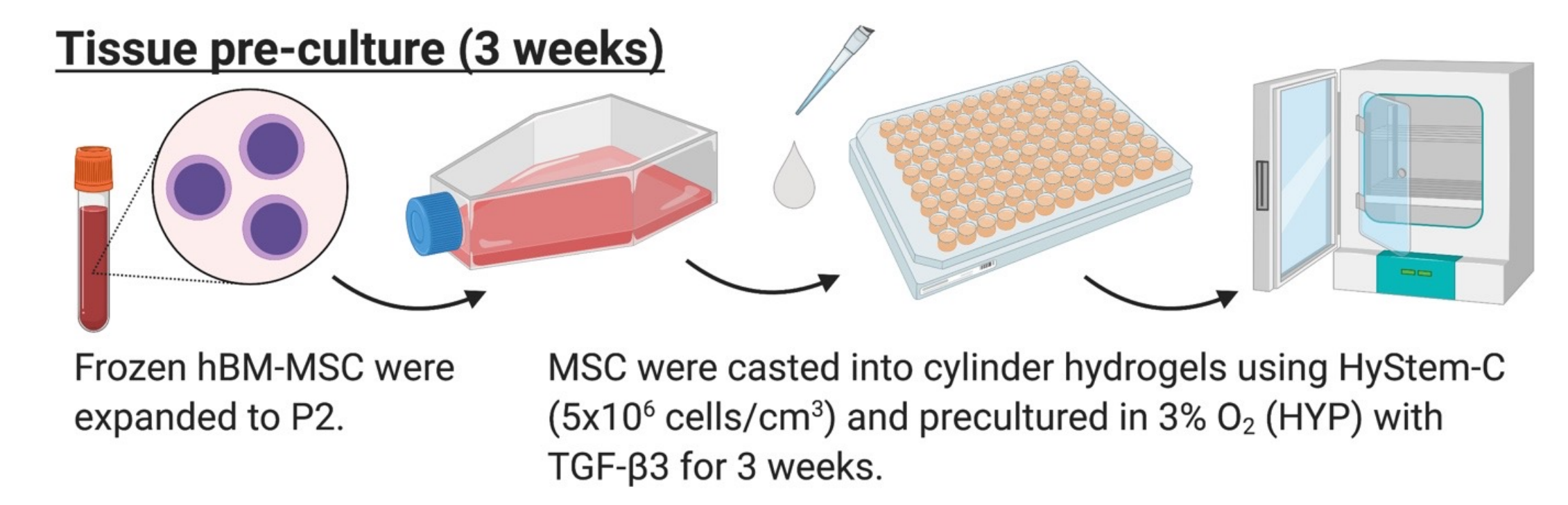
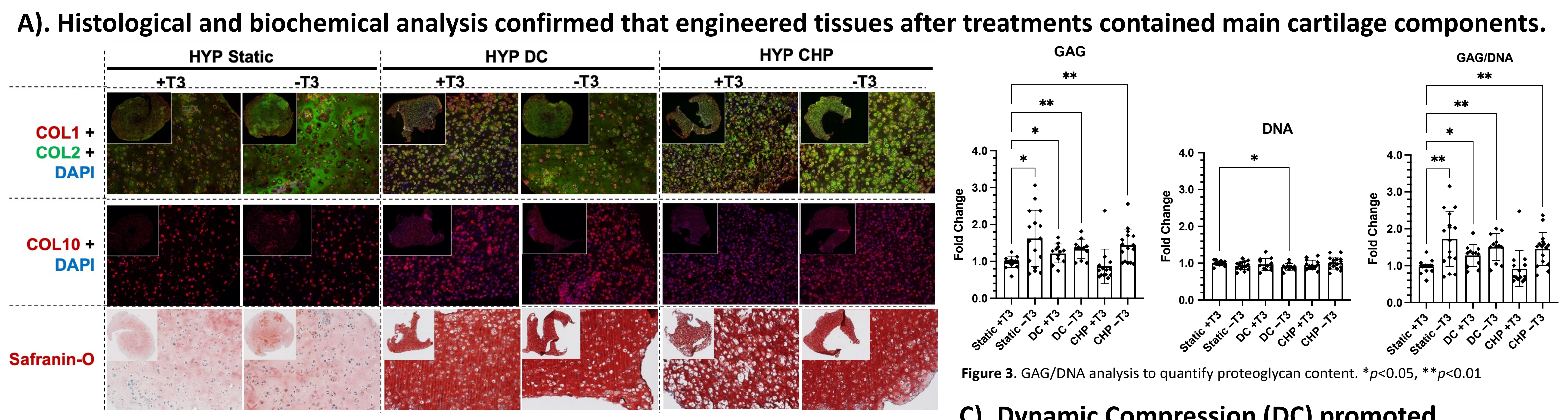


Figure 1. Experimental overview. Made with Biorender.com (2022)

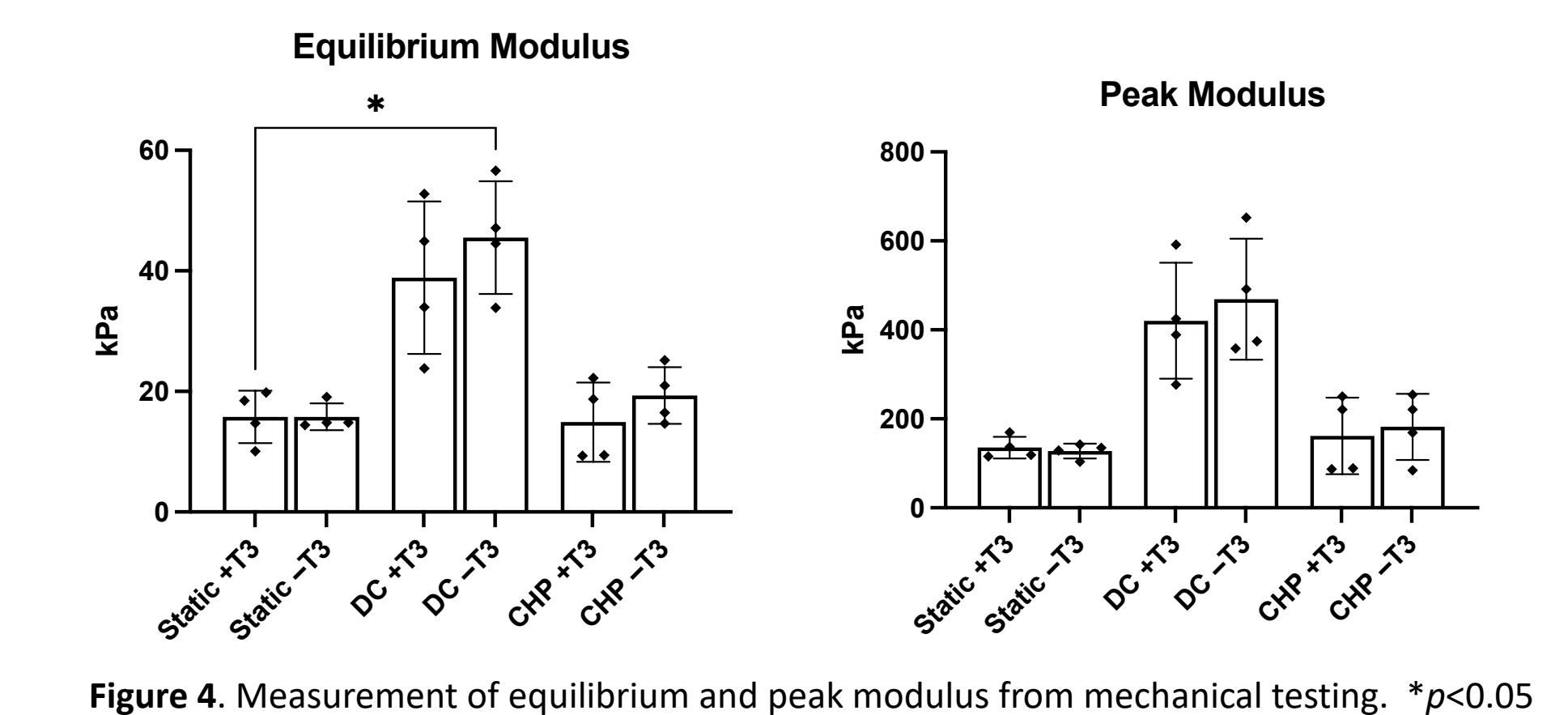
Results



B). RNA sequencing shows a significant (q < 0.05) modulation of select cartilage- and skeletogenesis-regulatory gene markers from treatments.

Gene	Description	Fold Changes	
		DC -T3 vs Static +T3	CHP -T3 vs Static +T3
<i>ASPN</i>	Asporin	13.38	5.61
<i>CHADL</i>	Chondroadherin Like	11.26	9.81
<i>CNMD</i>	Chondromodulin	71.43	54.94
<i>COL2A1</i>	Collagen Type II Alpha 1 Chain	7.78	6.99
<i>COL9A1</i>	Collagen Type IX Alpha 1 Chain	20.93	14.17
<i>COL9A2</i>	Collagen Type IX Alpha 2 Chain	7.10	5.92
<i>COL9A3</i>	Collagen Type IX Alpha 3 Chain	9.38	8.04
<i>COL11A2</i>	Collagen Type XI Alpha 2 Chain	12.17	9.62
<i>EPYC</i>	Epiphycan	13.44	7.69
<i>FRZB</i>	Frizzled Related Protein	23.91	42.05
<i>GDF10</i>	Growth Differentiation Factor 10	100.04	68.32
<i>GPM6B</i>	Glycoprotein M6B	10.52	10.98
<i>HAPLN1</i>	Hyaluronan And Proteoglycan Link Protein 1	6.24	6.26
<i>HAPLN3</i>	Hyaluronan And Proteoglycan Link Protein 3	-25.80	-7.15
<i>HAS2</i>	Hyaluronan Synthase 2	5.05	5.33
<i>MATN3</i>	Matrilin 3	9.48	9.68
<i>MATN4</i>	Matrilin 4	27.95	24.62
<i>MMP7</i>	Matrix Metalloproteinase 7	18.54	19.60
<i>MMP13</i>	Matrix Metalloproteinase 13	16.84	21.51
<i>PAPPA2</i>	Pappalysin 2	-6.68	-11.74
<i>PKD2</i>	Polycystin 2, Transient Receptor Potential Cation Channel	-41.85	-6.26
<i>PHOSPHO1</i>	Phosphoethanolamine/Phosphocholine Phosphatase 1	-28.54	-7.44
<i>PPARG</i>	Peroxisome Proliferator Activated Receptor Gamma	5.32	6.09
<i>PTX3</i>	Pentraxin 3	5.92	7.29
<i>SLC26A2</i>	Solute Carrier Family 26 Member 2	-20.12	-6.67
<i>SNORC</i>	Secondary Ossification Center Associated Regulator Of Chondrocyte Maturation	5.14	5.51
<i>SPARCL1</i>	SPARC Like 1	27.24	37.84
<i>TNC</i>	Tenascin C	6.91	7.69

C). Dynamic Compression (DC) promoted mechanical properties development



Conclusions

- Mechano-hypoxia conditioning with transient TGF-β3 stimulation promoted non-hypertrophic differentiation of hBM-MSC to chondrocytes.
- RNA seq data showed that the modality of loading treatment (DC vs CHP) impacts the level of gene expression which eventually consequents to different mechanical properties.
- The mechanical property development in the 3-week treatment shows promise toward generating functional engineered cartilage through more optimized and longer culture conditions.

Funding Sources

